



ARTICLE

Molecular Diagnostics

Urinary oestrogen steroidome as an indicator of the risk of localised prostate cancer progression

Jean-Philippe Emond¹, Louis Lacombe¹, Patrick Caron², Véronique Turcotte², David Simonyan³, Armen Aprikian⁴, Fred Saad⁵, Michel Carmel⁶, Simone Chevalier⁴, Chantal Guillemette² and Eric Lévesque¹

BACKGROUND: Prostate cancer (PCa) is the most common cancer in North American men. Beyond the established contribution of androgens to disease progression, growing evidence suggest that oestrogen-related pathways might also be of clinical importance. The aim of this study was to explore the association of urinary oestrogen levels with clinical outcomes.

METHODS: Urine samples from the prospective multi-institutional PROCURE cohort were collected before RP for discovery ($n = 259$) and validation ($n = 253$). Urinary total oestrogens (unconjugated + conjugated), including oestrone and oestradiol, their bioactive and inactive catechol and methyl derivatives ($n = 15$), were measured using mass spectrometry (MS).

RESULTS: The median follow-up time for the discovery and replication cohorts was 7.6 and 6.5 years, respectively. Highly significant correlations between urinary oestrogens were observed; however, correlations with circulating oestrogens were modest. Our findings indicate that higher levels of urinary oestriol and 16-ketoestradiol were associated with lower risk of BCR. In contrast, higher levels of 2-methoxyestrone were associated with an increased risk of development of metastasis/deaths.

CONCLUSIONS: Our data suggest that urinary levels of oestriol and 16-ketoestradiol metabolites are associated with a more favourable outcome, whereas those of 2-methoxyestrone are associated with an elevated risk of metastasis after RP. Further studies are required to better understand the impact of oestrogens on disease biology and as easily accessible urine-based risk-stratification markers.

British Journal of Cancer <https://doi.org/10.1038/s41416-021-01376-z>

BACKGROUND

Prostate cancer (PCa) remains the most frequent cancer in North American men¹ and the second most common cancer in men worldwide with over 1.2 million incident cases diagnosed in 2018 and over 350,000 deaths.² A recent study showed that the global burden of PCa is steadily increasing worldwide as well as the global incidence of the disease.³ It remains however a clinical challenge to predict the evolution of localised PCa at diagnosis. Clinical and pathological parameters bring important prognostic information, and scores like the CAPRA and CAPRA-S, were developed to predict the risk of BCR after radical prostatectomy (RP) in patients with localised disease.⁴ Despite this clinical and pathological information, additional patients' related factors are needed to better understand disease progression at early stages and to help anticipate the evolution of this heterogeneous disease.⁵ Knowing that ~25–30% of patients develop clinically evident recurrence in 15 years post RP,^{6–8} the incorporation of additional and non-invasive prognostic markers identifying these at-risk patients could further help clinicians and patients select on the best treatment and follow-up approaches to individualise care.

Most of the research on sex steroids was focused on androgens, such as testosterone (T) and dihydrotestosterone (DHT), and on the

androgen-receptor (AR) axis, which are known to play a major role in PCa pathophysiology, evolution and treatment strategies.^{9–14} However, growing evidence suggested that oestrogens might assist androgens in perpetuating carcinogenesis.^{15–21} Indeed, oestrogens are closely interrelated with androgens, notably due to the fact that T is the precursor of oestradiol (E_2). In animal models, when E_2 is combined with T, PCa incidence rises from 35 to 40% with androgen alone compared to 90–100% with E_2 .¹⁵ Moreover, *CYP19A1*- (aromatase) knockout mice, which lack oestrogen production, have elevated circulating T levels but they do not develop cancer.¹⁸ These observations are consistent with the fact that both types of hormones might be required for prostate carcinogenesis.^{16,19,20} In addition, the aromatase gene expression is altered in PCa,^{22,23} and polymorphisms in the *CYP19A1* and *ESR1* genes were both associated with progression in localised and advanced CRPC disease under androgen-deprivation therapy.^{24,25} More recently, aromatase expression in PCa cancer cells was associated with the development of metastasis through oestrogen-responsive elements in the *MMP-12* gene.²⁶ Besides, the potential role of downstream oestrogenic biotransformation pathways is supported by the demonstration of a link between oestrogen-metabolising genes such as *CYP1B1* and catechol-O-methyltransferase (*COMT*) and the risk of PCa.²⁷ Indeed,

¹Centre Hospitalier Universitaire (CHU) de Québec Research Center and Faculty of Medicine, Laval University, Québec, Canada; ²CHU de Québec Research Center and Faculty of Pharmacy, Laval University, Québec, Canada; ³Statistical and Clinical Research Platform, CHU de Québec Research Center, Québec, Canada; ⁴McGill University Health Center, McGill University, Faculty of Medicine, Québec, Canada; ⁵Centre Hospitalier de l'Université de Montréal, Université de Montréal, Québec, Canada and ⁶Université de Sherbrooke, Faculty of Medicine, Québec, Canada

Correspondence: Chantal Guillemette (chantal.guillemette@crchudequebec.ulaval.ca) or Eric Lévesque (eric.levesque@crchudequebec.ulaval.ca)

Received: 18 January 2021 Revised: 8 March 2021 Accepted: 16 March 2021

Published online: 07 April 2021

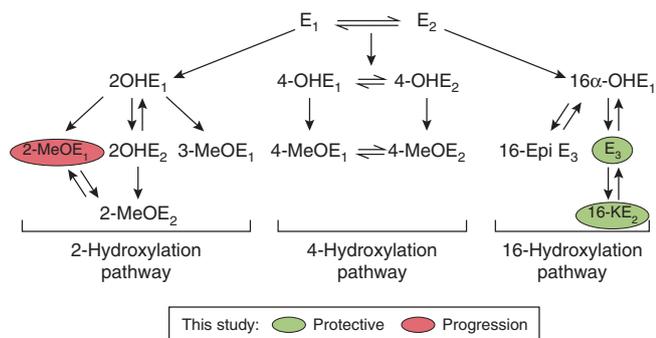


Fig. 1 Simplified biotransformation pathways for parent oestrogens, their bioactive and inactive catechol and methyl derivatives ($n = 15$) measured in this study. The associations of increasing levels of urinary oestrogens and clinical outcome observed in this study are highlighted in green (protective) and red (progression). E_1 : oestrone, E_2 : oestradiol, 2-OHE₁: 2-hydroxyestrone, 2-OHE₂: 2-hydroxyestradiol, 4-OHE₁: 4-hydroxyestrone, 4-OHE₂: 4-hydroxyestradiol, 16 α -OHE₁: 16 α -hydroxyestrone, E_3 : oestriol, 16-KE₂: 16-ketoestradiol, 16-epiE₃: 16-epiestriol, 2-MeOE₁: 2-methoxyestrone, 2-MeOE₂: 2-methoxyestradiol, 3-MeOE₁: 2-hydroxy-3-methoxyestrone, 4-MeOE₁: 4-methoxyestrone, 4-MeOE₂: 4-methoxyestradiol.

hydroxylation of oestrone (E_1) and E_2 by CYP enzymes can generate bioactive 2-hydroxy-catechol oestrogens (2-OHCE), 4-OHCE and 16-OHCE (Fig. 1). Unlike 2-OHCE, 4-OHCE and 16-OHCE retain oestrogenic activity and oestrogen-receptor (ER) binding affinities.^{28–31} Further, oxidation of CE derivatives to quinones can be deactivated by reduction with the quinone reductases' (NQO1 and NQO2) detoxification pathway or by covalently binding to DNA.^{32,33} The depurination of DNA adducts formed from 4-OHCE can generate mutations, whereas in contrast, 2-OHCE lacks carcinogenic potential.^{34–41} As a subsequent step, COMT can add a methyl group to CEs and the *O*-methylation process of 2-OHCE to 2-methoxy-CE (2-MeOCE) yields a metabolite that was shown to potentially inhibit cell proliferation, tubulin activity and angiogenesis.^{28,29,42–45} In addition, intraperitoneal infusion of 4-OHE₂ in animal models shows that the prostate region susceptible to neoplastic transformation has less protection by COMT and NQO detoxification pathways.⁴¹

Based on these findings, the purpose of this study was to evaluate whether parent oestrogen levels and their bioactive and inactive metabolites measured in urine of cancer patients are associated with adverse prognostic factors, biochemical recurrence (BCR) and development of metastasis/deaths in patients with localised PCa treated by prostatectomy. We used a mass spectrometry (MS) method with high sensitivity to measure levels of fifteen urinary oestrogens, including parent oestrogens, CEs and MeOCE to study a multi-institutional prospective PROCURE cohort⁴⁶ divided into discovery ($n = 259$) and replication/validation ($n = 253$) cohorts.

METHODS

Data from the prospective PROCURE PCa cohort

The multi-institutional prospective PROCURE biobank was previously described.^{47–49} Briefly, the overall cohort includes 2018 cases recruited between 2007 and 2012 at four university hospital centres in the Province of Québec in Canada (Montreal, McGill, Québec and Sherbrooke). All selected patients for this study had localised PCa at the time of diagnosis, underwent RP and did not receive androgen-deprivation therapy prior to urine sampling. For the discovery cohort, 259 patients were randomly selected in low- ($n = 51$), intermediate- ($n = 110$) and high-risk ($n = 98$) categories, providing an ~2:1 ratio with low-risk disease, to test their links with aggressive disease and adverse clinical outcomes such as BCR and metastasis. A predefined sample-size

selection composed of a higher percentage of high-risk patients was retained to detect a hazard ratio of 0.60 for BCR with a power of at least 80% at a 0.05 significance level with a standard deviation varying from 5 to 50. For the validation of positive findings, a cohort of 253 patients was also randomly selected from the same PROCURE cohort. Descriptive clinical and pathological characteristics of the patients studied from the discovery and validation PROCURE cohorts are shown in Table 1. The median follow-up time was 7.6 years for the discovery cohort and 6.5 years for the validation cohort. Serial prostate-specific antigen (PSA) measurements and clinical data were gathered during follow-up. All participants provided written informed consent and the CHU de Québec research ethics committee approved the research protocol (#2012-362). After prostatectomy, patients were seen regularly and PSA was measured every 3 months for two years, every 4–6 months for 2 years and then every 6–12 months or at the discretion of the physicians. BCR was defined as (1) the occurrence of a confirmed PSA > 0.2 ng/mL any time after surgery or (2) a detectable PSA of <0.2 ng/mL that triggered the initiation of salvage radiation or androgen-ablation therapy, as previously described.⁴⁶

Urinary oestrogen levels measured by mass spectrometry (MS) coupled to liquid chromatography

Urine samples were collected at a preoperative visit and centrifuged at 600 \times g for 10 min at 4 °C using standardised procedures and banked at –80 °C until analysis.⁴⁷ Plasma levels of steroids, including T, E_2 and E_1 , were available from a previous study of this cohort.⁴⁶ Fifteen oestrogens and their metabolites illustrated in Fig. 1 were extracted and quantified in urine samples using MS coupled to liquid chromatography, as previously described⁵⁰ and using a liquid chromatographic separation described by Franke et al.⁵¹ The approach utilises hydrolysis procedures to release the glucuronide and sulfate conjugates of each oestrogen and therefore represents the sum of unconjugated and conjugated quantified hormones.⁵⁰ The measured steroids in 250 μ L of urine were as follows: oestrone (E_1), oestradiol (E_2), oestriol (E_3), 16-epiestriol (16-epiE₃), 16-ketoestradiol (16-KE₂), 16 α -hydroxyestrone (16 α -OHE₁), 2-methoxyestrone (2-MeOE₁), 2-methoxyestradiol (2-MeOE₂), 2-hydroxyestrone (2-OHE₁), 2-hydroxyestradiol (2-OHE₂), 2-hydroxy-3-methoxyestrone (3-MeOE₁), 4-methoxyestrone (4-MeOE₁), 4-methoxyestradiol (4-MeOE₂), 4-hydroxyestrone (4-OHE₁) and 4-hydroxyestradiol (4-OHE₂). Reference steroids were purchased from Steraloids (Newport, RI, USA). Internal standards (deuterated steroids) were purchased from C/D/N Isotopes, Inc. (Pointe Claire, Québec, Canada) and were added to samples. Three low and high hormone-concentration quality-control replicates were included in each run, and all metabolite coefficients of variation were <10%. The limit of quantification, defined as the minimum value at which the ratio of signal-to-noise was $\geq 5:1$, for all these oestrogens, was 10 pg/mL in this biological fluid. Values of catechol oestrogens observed below the lower limit of quantification (LLOQ) were considered as undetected, even if detected above the limit of detection (LOD). All metabolites were detected in all patients, except 4-MeOE₂, 4-MeOE₁ and 4-OHE₂ that were above the LLOQ in 21%, 74% and 47% of the discovery cohort, respectively.

Urinary creatinine measurements

Normalisation of urinary oestrogen levels was achieved through urinary creatinine levels, measured by MS coupled to liquid chromatography, and reported as pg of steroid per mg of creatinine, in order to standardise the results and ensure comparability with previous studies.²⁹ Briefly, 20 μ L of urine (standard, QC or sample) were mixed with 5 mL of water and 100 μ L of internal standard (creatinine d3). Then 10 μ L of this dilution were mixed with 300 μ L of H₂O and injected into the MS. The chromatographic separation of creatinine was achieved using an Agilent 1200 (Agilent Technologies, Ville St-Laurent, QC, Canada). Separation was performed with an ACE Phenyl column

Table 1. Descriptive characteristics of studied patients from the PROCURE cohort.

Characteristics	Discovery cohort, n = 259	%	Validation cohort, n = 253	%
Mean age at diagnosis (yr)	62.2		63.1	
SD	6.1		6.3	
Range	45.4–78.7		47.0–76.5	
Median follow-up time (years)	7.6		6.5	
Biochemical recurrence (BCR)	127	49.0	139	54.9
Development of metastasis	19	7.4	24	9.5
Deaths	29	11.3	20	7.9
<i>PSA at diagnosis</i>				
≤10	212	81.9	200	79.1
>10 to 20	33	12.7	39	15.4
>20	14	5.4	14	5.5
<i>Pathologic Gleason score</i>				
≤6	61	23.6	54	21.3
7				
3 + 4	71	27.4	77	30.4
4 + 3	42	16.2	63	24.9
≥8	85	32.8	59	23.3
<i>Pathological T stage</i>				
≤pT2c	142	54.8	122	48.2
pT3a	81	31.3	68	26.9
≥pT3b	36	13.9	63	24.9
<i>Nodal invasion</i>				
pN ₀ or pN _x	242	93.4	230	90.1
pN ₁	17	6.6	23	9.1
<i>Margin status</i>				
Positive	114	44.0	132	52.2
Negative	145	56.0	121	47.8

(100 × 4.6-mm ID, particle size 3.0 µm, Canadian Life Science, Peterborough, Canada). A binary mobile phase, consisting of H₂O with 1 mM ammonium formate and 0.1% formic acid (solvent A), and MeOH with 1 mM ammonium formate and 0.1% formic acid (solvent B) was used at a flow rate of 0.9 mL/min. Analytes were eluted using the following program: 0–1 min, isocratic 10% B; 1–1.5 min, linear gradient 10–12.5% B; 1.5–1.6 min, linear gradient 12.5–90% B; 1.6–2.6 min, isocratic 90% B; 2.6–2.7 min, linear gradient 90–10% B; 2.7–6.0 min, isocratic 10% B. The API 3200 triple-quadrupole MS instrument (AB Sciex, Concord, ON, Canada) was equipped with a turbo ion-spray source, configured in multiple reaction-monitoring (MRM) mode. MRM transitions were 114.3→44.1 for creatinine and 117.2→47.2 for creatinine d₃. Electrospray ionisation was performed in the positive mode with an ionisation voltage of 5000 V, a declustering potential of 36 V, collision energy of 27 V and a heater probe temperature at 500 °C. Analyst software version 1.6.2 (AB Sciex, Concord, ON, Canada) was used for system control and data analysis.

Statistical analysis

Quantitative variables were described as mean, standard deviation (SD), range, median and interquartile range (Q1;Q3), and

qualitative variables as frequencies and percentages. Spearman correlations (r_s) between urinary oestrogens were obtained and tested for H₀: $r_s = 0$. The associations between clinical prognostic parameters (PSA, Gleason score, pT) and natural logarithm-transformed urinary oestrogen levels were estimated using generalised linear regression modelling adjusted for age and body mass index (BMI). The log transformation of urinary oestrogens was used due to the non-normal distribution of the variables as performed in studies similar in scope.^{52,53} A weighted multivariable proportional hazard Cox model adjusted for age, surgical margin, pN, PSA (continuous variable), pT (<pT3a, pT3a, and >pT3a) and Gleason score (GS 6, GS3 + 4, GS4 + 3 and GS > 7) was used to evaluate the risk of BCR and metastasis/deaths according to urinary oestrogen levels. BMI was included in the final multivariable model as a continuous variable. A *P* value of < 0.05 was considered statistically significant. Based on the fact that (i) all tested hormones are positioned in the same metabolic pathway and (ii) we validated findings in a separate dataset, no correction for multiple testing was performed since no random hypotheses were tested herein. The software used was SAS 9.4 by SAS Institute Inc. (Cary, NC).

RESULTS

Correlations between urinary oestrogens and their metabolites
 The parent oestrogen E₁ and its metabolites E₃ and 2-OHE₁ were the most abundant urinary oestrogens found in patients at 4875.6 pg/mg creatinine (95% CI: 4566.3–5184.9), 4852.6 pg/mg creatinine (95% CI: 4470.1–5235.1) and 2810.7 pg/mg creatinine (95% CI: 2573.8–3047.7) (Supplementary Table 1). E₂ was the fourth most abundant oestrogen measured in urine with 1318.5 pg/mg creatinine (95% CI: 1205.1–1431.9). Spearman correlations revealed that urinary parent oestrogens E₁ and E₂ strongly correlated ($r_s = 0.76$, $P < 0.001$) with each other and also with the levels of 2-OHE₁, 2-OHE₂, 16-KE₂ and 16-epiE₃ ($P < 0.001$) (Fig. 2).

Levels of urinary oestrogens and their correlation to circulating levels of parent oestrogens and testosterone (T)

Moderate correlations were observed between circulating T, E₂ and E₁ and urinary oestrogen levels, with the most significant being with E₂ in urine. For example, urinary E₂ correlated with plasma E₂ with a Spearman correlation value of $r_s = 0.133$ ($P = 0.032$). The strongest correlations were plasma E₂ with 16-KE₂ ($r_s = 0.246$, $P < 0.0001$) and E₃ ($r_s = 0.230$, $P = 0.0002$). Plasma T levels correlated to 16-KE₂ ($r_s = 0.152$, $P = 0.014$). All correlations are presented in Supplementary Table 2.

Relationships between established prognostic factors and urinary oestrogen levels

No significant associations between urinary oestrogens and PSA levels were observed (Supplementary Table 3). In the discovery cohort, 4-MeOE₁ and 4-OHE₁ urinary levels were linked to Gleason scores (Supplementary Table 4), but these two associations were not maintained in the validation cohort (not shown). Urinary creatinine levels were not associated with Gleason scores ($P = 0.947$). Urinary oestrogen levels were not associated with pT staging (Supplementary Table 5). Both 2-MeOE₁ and 4-OHE₂ were associated with changes in body mass index (BMI) (Supplementary Table 6).

Urinary oestrogen levels and the risk of progression after RP

As shown previously for the entire PROCURE cohort,⁴⁹ BMI was not associated with clinical outcome herein (hazard ratio (HR) = 0.98, 95% CI: 0.93–1.05, $P = 0.625$). In multivariable analyses, increasing levels of urinary E₃ were associated with a reduced risk of BCR with HR values of 0.66 (95% CI: 0.43–0.98, $P = 0.038$) in the discovery cohort and 0.74 (95% CI: 0.52–1.04, $P = 0.086$) in the validation cohort. A HR of 0.74 (95% CI: 0.60–0.91, $P = 0.006$) is reported for the combined cohorts. Increased levels of the oestradiol

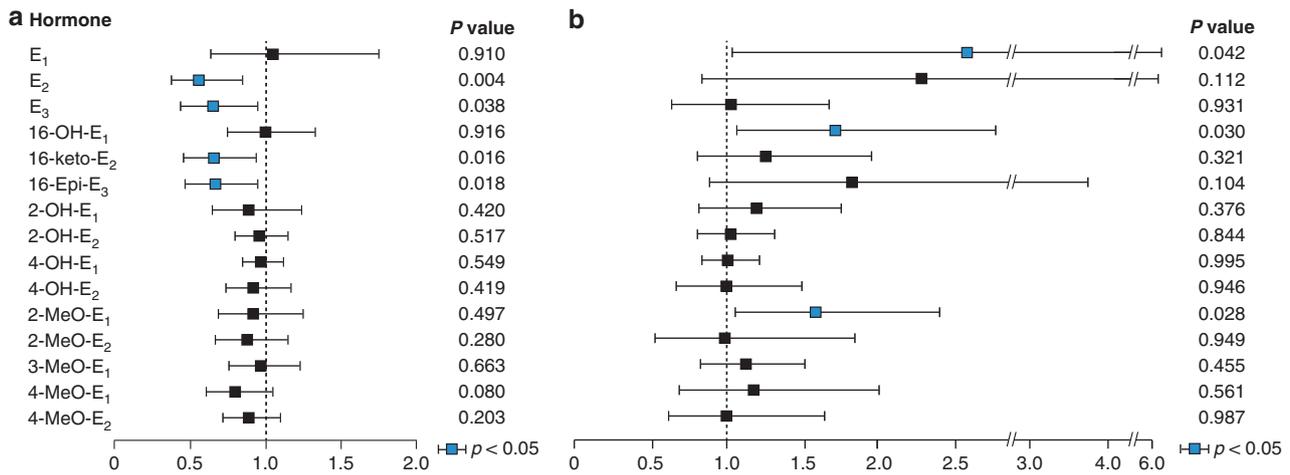


Fig. 3 Urinary oestrogens and the risk of PCa progression. Association between total urinary oestrogens and the risk of (a) BCR and (b) metastasis/deaths in men with localised PCa in the discovery cohort ($n = 259$). Boxes shown within the main panels represent hazard ratios (HRs) and their 95% confidence intervals in multivariable analyses adjusted for age, PSA, Gleason score, surgical margin, pT, pN and BMI for each hormone. Each unit increment in log-transformed hormone levels is associated with the indicated changes in HR values.

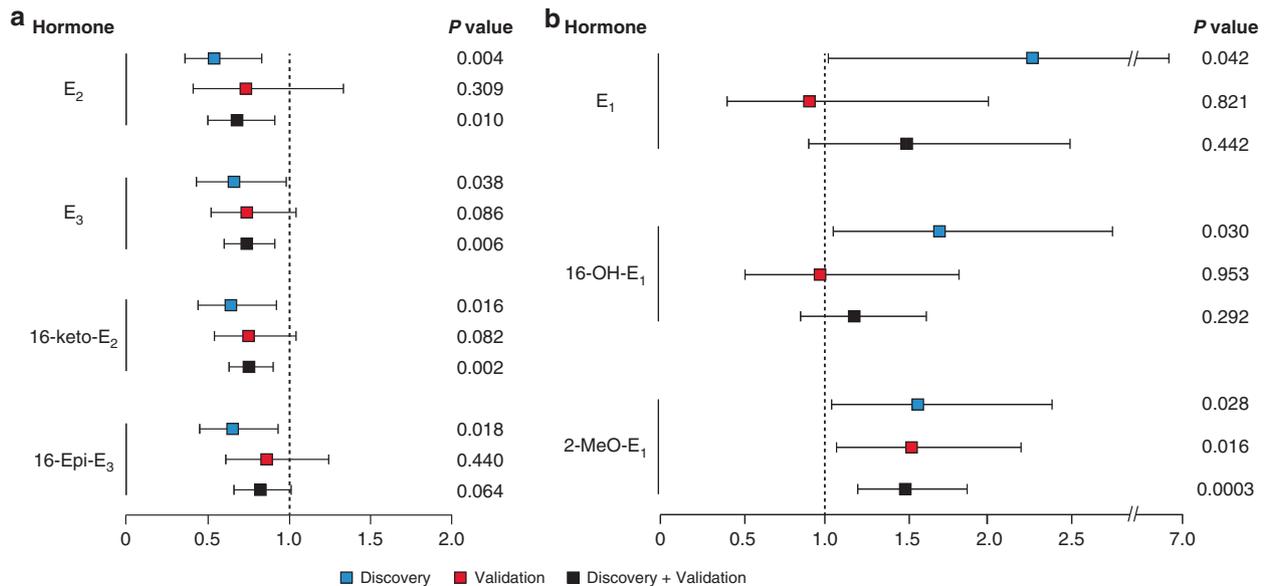


Fig. 4 Validation of urinary oestrogens and the risk of PCa progression. Validation of positive associations between total urinary oestrogens and the risk of (a) BCR and (b) metastasis/deaths in men with localised PCa in a confirmatory cohort of 253 patients originating from PROCURE. Boxes shown within the main panels represent hazard ratios (HRs) and their 95% confidence intervals in multivariable analyses adjusted for age, PSA, Gleason score, surgical margin, pT, pN and BMI for each hormone. Each unit increment in log-transformed hormone levels is associated with the indicated changes in HR values.

conjugated (sulfate and glucuronide) forms and (3) inaccessibility of intraprostatic oestrogen levels.

We conclude that urinary E₃ and 16-KE₂ levels are associated with less BCR events after RP, and 2-MeOE₁ was associated with disease progression. Our observations are in support of a potential role of oestrogens in progression, but require further evaluation in additional independent cohorts to confirm the associations observed. Our data raise the possibility that an assessment of the oestrogen metabolome in urine samples may provide clinically relevant information that can be used alone or combined with circulating, tumoral and germline markers to better predict clinical outcomes of PCa patients at the time of diagnosis. Additional studies are definitely required to evaluate if urinary oestrogens can be used along with the circulating steroidome to help predict

disease evolution at the time of initial diagnosis. Future studies aimed at understanding the impact of the complex oestrogen pathway in PCa are undeniably warranted.

ACKNOWLEDGEMENTS

The authors are thankful to all participating patients and staff at each site associated with the PROCURE Biobank who have made this scientific contribution possible.

AUTHOR CONTRIBUTIONS

Study concept, design and supervision: E.L. and C.G. Steroid measurements: P.C., V.T. and C.G. Statistical analysis: D.S. Interpretation of the data: J.P.E., L.L., P.C., V.T., D.S., A.A., F.S., M.C., S.C., C.G. and E.L. Drafting of the paper: J.P.E., C.G. and E.L. Critical

revision of the paper for important intellectual content: J.P.E., L.L., P.C., V.T., D.S., A.A., F.S., M.C., S.C., C.G. and E.L. Patients' recruitment and clinical data: F.S., L.L., M.C., S.C. and A.A. Obtaining funding: E.L., C.G. and L.L.

ADDITIONAL INFORMATION

Ethics approval and consent to participate All participants provided written informed consent and the CHU de Quebec research ethics committee approved the research protocol, and the study was performed in accordance with the Declaration of Helsinki.

Consent to publish Not applicable.

Data availability The datasets analysed during this study are available from the corresponding author on reasonable request.

Competing interests The authors declare no competing interests.

Funding information This work was supported by research grants from Prostate Cancer Canada (D2013-22 to E.L., C.G. and L.L.), the Cancer Research Society (to C.G.) and the Canada Research Chair Program (to C.G.). E.L. holds a CIHR Clinician-Scientist Award and was the holder of the PCC rising star award (RS2013-59). C.G. holds the Canada Research Chair in Pharmacogenomics (Tier I). The results are based on samples and patient data obtained from the PROCURE Biobank, supported by donations in a partnership with the Cancer Research Society of Canada.

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41416-021-01376-z>.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

REFERENCES

1. Miller, K. D., Nogueira, L., Mariotto, A. B., Rowland, J. H., Yabroff, K. R., Alfano, C. M. et al. Cancer treatment and survivorship statistics, 2019. *CA Cancer J. Clin.* **69**, 363–385 (2019).
2. Bray, F., Ferlay, J., Soerjomataram, I., Siegel, R. L., Torre, L. A. & Jemal, A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* **68**, 394–424 (2018).
3. Pishgar, F., Ebrahimi, H., Saeedi Moghaddam, S., Fitzmaurice, C. & Amini, E. Global, regional and national burden of prostate cancer, 1990 to 2015: results from the Global Burden of Disease Study 2015. *J. Urol.* **199**, 1224–1232 (2018).
4. Lorent, M., Maalmi, H., Tessier, P., Supiot, S., Dantan, E. & Foucher, Y. Meta-analysis of predictive models to assess the clinical validity and utility for patient-centered medical decision making: application to the cancer of the prostate risk assessment (CAPRA). *BMC Med. Inf. Decis. Mak.* **19**, 2 (2019).
5. Fraser, M., Sabelykova, V. Y., Yamaguchi, T. N., Heisler, L. E., Livingstone, J., Huang, V. et al. Genomic hallmarks of localized, non-indolent prostate cancer. *Nature* **541**, 359–364 (2017).
6. Pound, C. R., Partin, A. W., Eisenberger, M. A., Chan, D. W., Pearson, J. D. & Walsh, P. C. Natural history of progression after PSA elevation following radical prostatectomy. *J. Am. Med. Assoc.* **281**, 1591–1597 (1999).
7. Audet-Walsh, E., Bellemare, J., Lacombe, L., Fradet, Y., Fradet, V., Douville, P. et al. The impact of germline genetic variations in hydroxysteroid (17-beta) dehydrogenases on prostate cancer outcomes after prostatectomy. *Eur. Urol.* **62**, 88–96 (2012).
8. Audet-Walsh, E., Bellemare, J., Nadeau, G., Lacombe, L., Fradet, Y., Fradet, V. et al. SRD5A polymorphisms and biochemical failure after radical prostatectomy. *Eur. Urol.* **60**, 1226–1234 (2011).
9. Huggins, C. & Clark, P. J. Quantitative studies of prostatic secretion: II. the effect of castration and of estrogen injection on the normal and on the hyperplastic prostate glands of dogs. *J. Exp. Med.* **72**, 747–762 (1940).
10. Schileyko, A. A. & Solovyeva, E. N. On the taxonomic position of the enigmatic genus *Tonkinodentus* Schileyko, 1992 (Chilopoda, Scolopendromorpha): the first molecular data. *Zookeys* **840**, 133–155 (2019).
11. Ryan, C. J., Smith, M. R., de Bono, J. S., Molina, A., Logothetis, C. J., de Souza, P. et al. Abiraterone in metastatic prostate cancer without previous chemotherapy. *N. Engl. J. Med.* **368**, 138–148 (2013).
12. Ryan, C. J., Smith, M. R., Fizazi, K., Saad, F., Mulders, P. F., Sternberg, C. N. et al. Abiraterone acetate plus prednisone versus placebo plus prednisone in chemotherapy-naïve men with metastatic castration-resistant prostate cancer

(COU-AA-302): final overall survival analysis of a randomised, double-blind, placebo-controlled phase 3 study. *Lancet Oncol.* **16**, 152–160 (2015).

13. Beer, T. M., Armstrong, A. J., Rathkopf, D., Loriot, Y., Sternberg, C. N., Higano, C. S. et al. Enzalutamide in men with chemotherapy-naïve metastatic castration-resistant prostate cancer: extended analysis of the phase 3 PREVAIL study. *Eur. Urol.* **71**, 151–154 (2017).
14. Scher, H. I., Fizazi, K., Saad, F., Taplin, M. E., Sternberg, C. N., Miller, K. et al. Increased survival with enzalutamide in prostate cancer after chemotherapy. *N. Engl. J. Med.* **367**, 1187–1197 (2012).
15. Bosland, M. C., Ford, H. & Horton, L. Induction at high incidence of ductal prostate adenocarcinomas in NBL/Cr and Sprague-Dawley Hsd:SD rats treated with a combination of testosterone and estradiol-17 beta or diethylstilbestrol. *Carcinogenesis* **16**, 1311–1317 (1995).
16. Bosland, M. C. & Mahmoud, A. M. Hormones and prostate carcinogenesis: androgens and estrogens. *J. Carcinog.* **10**, 33 (2011).
17. McPherson, S. J., Ellem, S. J. & Risbridger, G. P. Estrogen-regulated development and differentiation of the prostate. *Differentiation* **76**, 660–670 (2008).
18. McPherson, S. J., Wang, H., Jones, M. E., Pedersen, J., Iismaa, T. P., Wreford, N. et al. Elevated androgens and prolactin in aromatase-deficient mice cause enlargement, but not malignancy, of the prostate gland. *Endocrinology* **142**, 2458–2467 (2001).
19. Weng, C., Cai, J., Wen, J., Yuan, H., Yang, K., Imperato-McGinley, J. et al. Differential effects of estrogen receptor ligands on regulation of dihydrotestosterone-induced cell proliferation in endothelial and prostate cancer cells. *Int. J. Oncol.* **42**, 327–337 (2013).
20. Yu, L., Shi, J., Cheng, S., Zhu, Y., Zhao, X., Yang, K. et al. Estrogen promotes prostate cancer cell migration via paracrine release of ENO1 from stromal cells. *Mol. Endocrinol.* **26**, 1521–1530 (2012).
21. Yang, L., Gaikwad, N. W., Meza, J., Cavalieri, E. L., Muti, P., Trock, B. et al. Novel biomarkers for risk of prostate cancer: results from a case-control study. *Prostate* **69**, 41–48 (2009).
22. Ellem, S. J., Schmitt, J. F., Pedersen, J. S., Frydenberg, M. & Risbridger, G. P. Local aromatase expression in human prostate is altered in malignancy. *J. Clin. Endocrinol. Metab.* **89**, 2434–2441 (2004).
23. Montgomery, R. B., Mostaghel, E. A., Vessella, R., Hess, D. L., Kalhorn, T. F., Higano, C. S. et al. Maintenance of intratumoral androgens in metastatic prostate cancer: a mechanism for castration-resistant tumor growth. *Cancer Res.* **68**, 4447–4454 (2008).
24. Ross, R. W., Oh, W. K., Xie, W., Pomerantz, M., Nakabayashi, M., Sartor, O. et al. Inherited variation in the androgen pathway is associated with the efficacy of androgen-deprivation therapy in men with prostate cancer. *J. Clin. Oncol.* **26**, 842–847 (2008).
25. Adams, M. J., Johnson, S. A., Lefevre, P., Levesque, V., Hayward, V., Andre, T. et al. Finger pad friction and its role in grip and touch. *J. R. Soc. Interface* **10**, 20120467 (2013).
26. Duan, L., Chen, Z., Lu, J., Liang, Y., Wang, M., Roggero, C. M. et al. Histone lysine demethylase KDM4B regulates the alternative splicing of the androgen receptor in response to androgen deprivation. *Nucleic Acids Res.* **47**, 11623–11636 (2019).
27. Cussenot, O., Azzouzi, A. R., Nicolaiew, N., Fromont, G., Mangin, P., Cormier, L. et al. Combination of polymorphisms from genes related to estrogen metabolism and risk of prostate cancers: the hidden face of estrogens. *J. Clin. Oncol.* **25**, 3596–3602 (2007).
28. Zhu, B. T., Han, G. Z., Shim, J. Y., Wen, Y. & Jiang, X. R. Quantitative structure-activity relationship of various endogenous estrogen metabolites for human estrogen receptor alpha and beta subtypes: Insights into the structural determinants favoring a differential subtype binding. *Endocrinology* **147**, 4132–4150 (2006).
29. Zhu, B. T. & Conney, A. H. Is 2-methoxyestradiol an endogenous estrogen metabolite that inhibits mammary carcinogenesis? *Cancer Res.* **58**, 2269–2277 (1998).
30. Zhu, B. T. & Conney, A. H. Functional role of estrogen metabolism in target cells: review and perspectives. *Carcinogenesis* **19**, 1–27 (1998).
31. Han, D. F., Zhou, X., Hu, M. B., Xie, W., Mao, Z. F., Chen, D. E. et al. Polymorphisms of estrogen-metabolizing genes and breast cancer risk: a multigenic study. *Chin. Med. J.* **118**, 1507–1516 (2005).
32. Cavalieri, E. L. & Rogan, E. G. Unbalanced metabolism of endogenous estrogens in the etiology and prevention of human cancer. *J. Steroid Biochem. Mol. Biol.* **125**, 169–180 (2011).
33. Lee, M. M., Gomez, S. L., Chang, J. S., Wey, M., Wang, R. T. & Hsing, A. W. Soy and isoflavone consumption in relation to prostate cancer risk in China. *Cancer Epidemiol. Biomark. Prev.* **12**, 665–668 (2003).
34. Saeed, M., Rogan, E. & Cavalieri, E. Mechanism of metabolic activation and DNA adduct formation by the human carcinogen diethylstilbestrol: the defining link to natural estrogens. *Int. J. Cancer* **124**, 1276–1284 (2009).

35. Cavalieri, E. & Rogan, E. Catechol quinones of estrogens in the initiation of breast, prostate, and other human cancers: keynote lecture. *Ann. N. Y. Acad. Sci.* **1089**, 286–301 (2006).
36. Saeed, M., Gunselman, S. J., Higginbotham, S., Rogan, E. & Cavalieri, E. Formation of the depurinating N3adenine and N7guanine adducts by reaction of DNA with hexestrol-3',4'-quinone or enzyme-activated 3'-hydroxyhexestrol. Implications for a unifying mechanism of tumor initiation by natural and synthetic estrogens. *Steroids* **70**, 37–45 (2005).
37. Cavalieri, E., Frenkel, K., Liehr, J. G., Rogan, E. & Roy, D. Estrogens as endogenous genotoxic agents—DNA adducts and mutations. *J. Natl. Cancer Inst. Monogr.* **27**, 75–93 (2000).
38. Sissung, T. M., Danesi, R., Price, D. K., Steinberg, S. M., de Wit, R., Zahid, M. et al. Association of the CYP1B1*3 allele with survival in patients with prostate cancer receiving docetaxel. *Mol. Cancer Ther.* **7**, 19–26 (2008).
39. Markushin, Y., Gaikwad, N., Zhang, H., Kapke, P., Rogan, E. G., Cavalieri, E. L. et al. Potential biomarker for early risk assessment of prostate cancer. *Prostate* **66**, 1565–1571 (2006).
40. Rogan, E. G. & Cavalieri, E. L. Estrogen metabolites, conjugates, and DNA adducts: possible biomarkers for risk of breast, prostate, and other human cancers. *Adv. Clin. Chem.* **38**, 135–149 (2004).
41. Cavalieri, E. L., Devanesan, P., Bosland, M. C., Badawi, A. F. & Rogan, E. G. Catechol estrogen metabolites and conjugates in different regions of the prostate of Noble rats treated with 4-hydroxyestradiol: implications for estrogen-induced initiation of prostate cancer. *Carcinogenesis* **23**, 329–333 (2002).
42. D'Amato, R. J., Lin, C. M., Flynn, E., Folkman, J. & Hamel, E. 2-Methoxyestradiol, an endogenous mammalian metabolite, inhibits tubulin polymerization by interacting at the colchicine site. *Proc. Natl Acad. Sci. USA* **91**, 3964–3968 (1994).
43. Fotsis, T., Zhang, Y., Pepper, M. S., Adlercreutz, H., Montesano, R., Nawroth, P. P. et al. The endogenous oestrogen metabolite 2-methoxyoestradiol inhibits angiogenesis and suppresses tumour growth. *Nature* **368**, 237–239 (1994).
44. Lakhani, N. J., Sarkar, M. A., Venitz, J. & Figg, W. D. 2-Methoxyestradiol, a promising anticancer agent. *Pharmacotherapy* **23**, 165–172 (2003).
45. Zhu, B. T. & Liehr, J. G. Inhibition of the catechol-O-methyltransferase-catalyzed O-methylation of 2- and 4-hydroxyestradiol by catecholamine: implications for the mechanism of estrogen-induced carcinogenesis. *Arch. Biochem. Biophys.* **304**, 248–256 (1993).
46. Lévesque, E., Caron, P., Lacombe, L., Turcotte, V., Simonyan, D., Fradet, Y. et al. comprehensive analysis of steroid hormones and progression of localized high-risk prostate cancer. *Cancer Epidemiol. Biomark. Prev.* **28**, 701–706 (2019).
47. Brimo, F., Aprikian, A., Latour, M., Tetu, B., Doueik, A., Scarlata, E. et al. Strategies for biochemical and pathologic quality assurance in a large multi-institutional biorepository; the experience of the PROCURE Quebec Prostate Cancer Biobank. *Biopreserv. Biobank.* **11**, 285–290 (2013).
48. Levesque, E., Caron, P., Lacombe, L., Turcotte, V., Simonyan, D., Fradet, Y. et al. A comprehensive analysis of steroid hormones and progression of localized high-risk prostate cancer. *Cancer Epidemiol. Biomark. Prev.* **28**, 701–706 (2019).
49. Wissing, M., Chevalier, S., McKercher, G., Laprise, C., Aprikian, S., O'Flaherty, A. et al. The relationship between body-mass index, physical activity, and pathologic and clinical outcomes after radical prostatectomy for prostate cancer. *World J. Urol.* **37**, 789–798 (2019).
50. Audet-Delage, Y., Gregoire, J., Caron, P., Turcotte, V., Plante, M., Ayotte, P. et al. Estradiol metabolites as biomarkers of endometrial cancer prognosis after surgery. *J. Steroid Biochem. Mol. Biol.* **178**, 45–54 (2018).
51. Franke, A. A., Custer, L. J., Morimoto, Y., Nordt, F. J. & Maskarinec, G. Analysis of urinary estrogens, their oxidized metabolites, and other endogenous steroids by benchtop orbitrap LCMS versus traditional quadrupole GCMS. *Anal. Bioanal. Chem.* **401**, 1319–1330 (2011).
52. Petrick, J. L., Hyland, P. L., Caron, P., Falk, R. T., Pfeiffer, R. M., Dawsey, S. M. et al. Associations between prediagnostic concentrations of circulating sex steroid hormones and esophageal/gastric cardia adenocarcinoma among men. *J. Natl Cancer Inst.* **111**, 34–41 (2019).
53. Eliassen, A. H., Missmer, S. A., Tworoger, S. S. & Hankinson, S. E. Endogenous steroid hormone concentrations and risk of breast cancer: does the association vary by a woman's predicted breast cancer risk? *J. Clin. Oncol.* **24**, 1823–1830 (2006).
54. Dobbs, R. W., Malhotra, N. R., Greenwald, D. T., Wang, A. Y., Prins, G. S. & Abern, M. R. Estrogens and prostate cancer. *Prostate Cancer Prostatic Dis.* **22**, 59–65 (2019).
55. Cavalieri, E. L. & Rogan, E. G. Depurinating estrogen-DNA adducts, generators of cancer initiation: their minimization leads to cancer prevention. *Clin. Transl. Med.* **5**, 12 (2016).
56. Toren, P., Hoffman, A., Ding, K., Joncas, F. H., Turcotte, V., Caron, P. et al. Serum sex steroids as prognostic biomarkers in patients receiving androgen deprivation therapy for recurrent prostate cancer: a post hoc analysis of the PR.7 trial. *Clin. Cancer Res.* **24**, 5305–5312 (2018).
57. Di Zazzo, E., Galasso, G., Giovannelli, P., Di Donato, M. & Castoria, G. Estrogens and their receptors in prostate cancer: therapeutic implications. *Front. Oncol.* **8**, 2 (2018).
58. Levesque, E., Laverdiere, I., Audet-Walsh, E., Caron, P., Rouleau, M., Fradet, Y. et al. Steroidogenic germline polymorphism predictors of prostate cancer progression in the estradiol pathway. *Clin. Cancer Res.* **20**, 2971–2983 (2014).
59. Salonia, A., Abdollah, F., Capitanio, U., Suardi, N., Briganti, A., Gallina, A. et al. Serum sex steroids depict a nonlinear u-shaped association with high-risk prostate cancer at radical prostatectomy. *Clin. Cancer Res.* **18**, 3648–3657 (2012).
60. Muti, P., Westerlind, K., Wu, T., Grimaldi, T., De Berry, J. 3rd, Schunemann, H. et al. Urinary estrogen metabolites and prostate cancer: a case-control study in the United States. *Cancer Causes Control* **13**, 947–955 (2002).
61. Barba, M., Yang, L., Schunemann, H. J., Sperati, F., Grioni, S., Stranges, S. et al. Urinary estrogen metabolites and prostate cancer: a case-control study and meta-analysis. *J. Exp. Clin. Cancer Res.* **28**, 135 (2009).
62. Teas, J., Cunningham, J. E., Fowke, J. H., Nitcheva, D., Kanwat, C. P., Boulware, R. J. et al. Urinary estrogen metabolites, prostate specific antigen, and body mass index among African-American men in South Carolina. *Cancer Detect. Prev.* **29**, 494–500 (2005).
63. Kostı, O., Xu, X., Veenstra, T. D., Hsing, A. W., Chu, L. W., Goldman, L. et al. Urinary estrogen metabolites and prostate cancer risk: a pilot study. *Prostate* **71**, 507–516 (2011).
64. Schumacher, G. & Neuhaus, P. The physiological estrogen metabolite 2-methoxyestradiol reduces tumor growth and induces apoptosis in human solid tumors. *J. Cancer Res. Clin. Oncol.* **127**, 405–410 (2001).
65. Alfaro, A., Leon, A., Guajardo-Correa, E., Reuquen, P., Torres, F., Mery, M. et al. MgO nanoparticles coated with polyethylene glycol as carrier for 2-Methoxyestradiol anticancer drug. *PLoS ONE* **14**, e0214900 (2019).